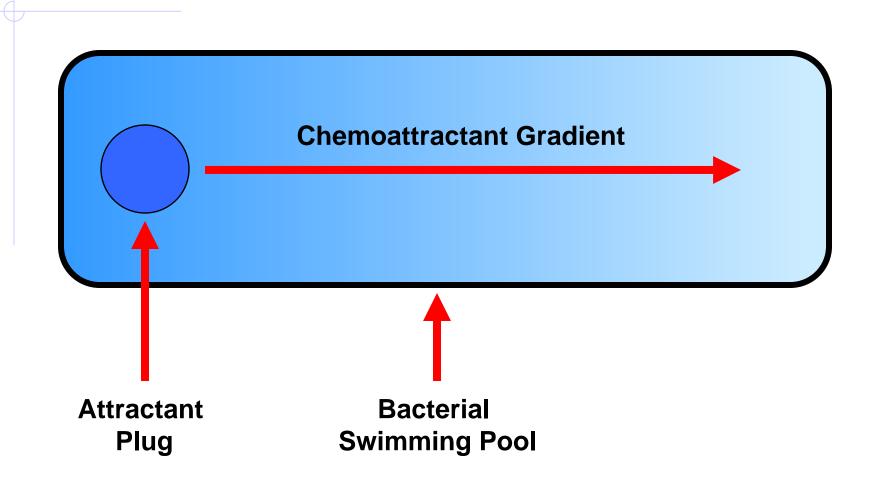
Synchronized Chemotactic Oscillators

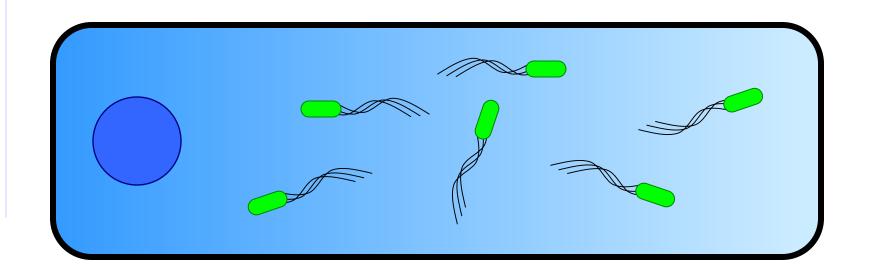
S.M.U.G.

Summer Synthetic Biology Competition Massachusetts Institute of Technology November 6, 2004

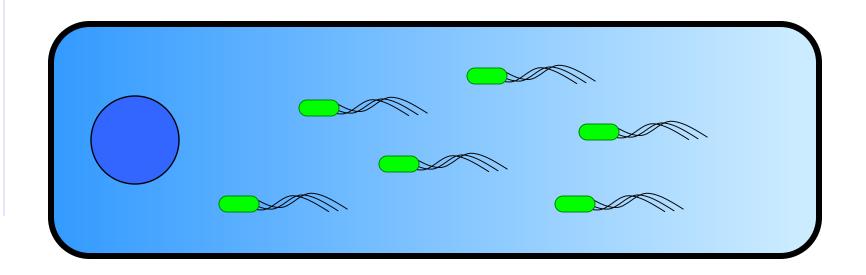
Motivation

- Our goal: an interesting, complex system something cool. But how to make it happen?
- We focused on implementing modularity
 - Breaking biological systems into modular pieces
 - At a low level, this is BioBricks
- Building a modular system allowed efficient division of labor – key for a team this large



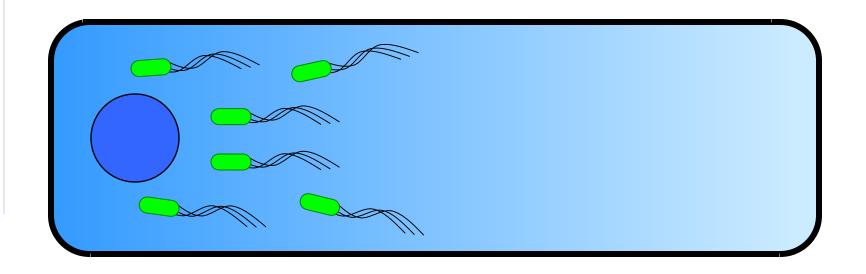


Bacteria are added to the swimming pool

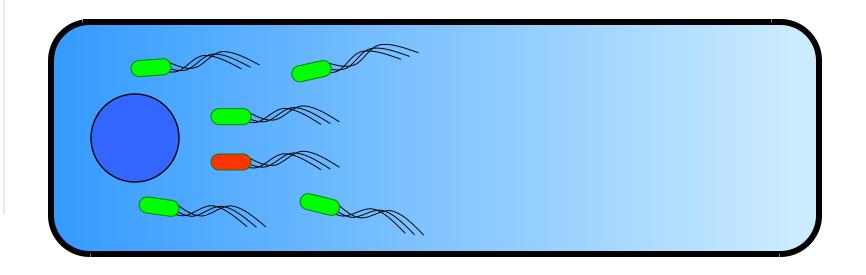


Chemotaxis is enabled, indicated by green bacteria

Bacteria start swimming up the gradient towards the attractant plug

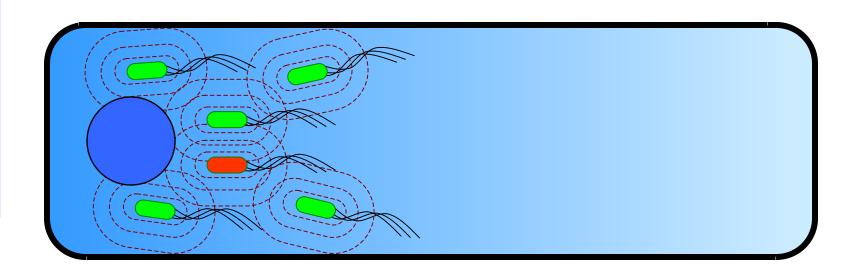


The bacteria congregate around the attractant plug

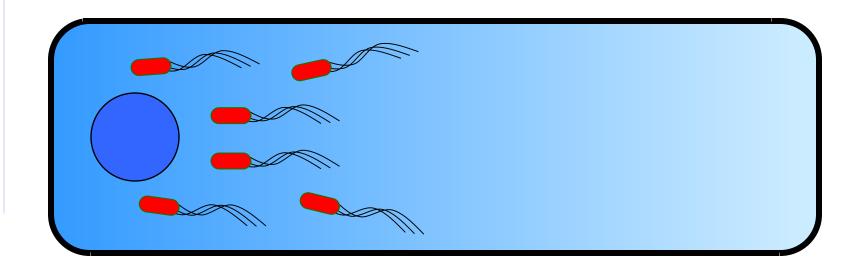


Each bacteria has an internal oscillator, driving switch between:

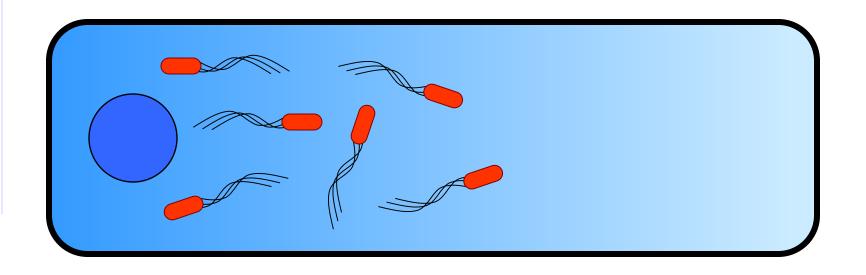
chemotaxis enabled chemotaxis disabled



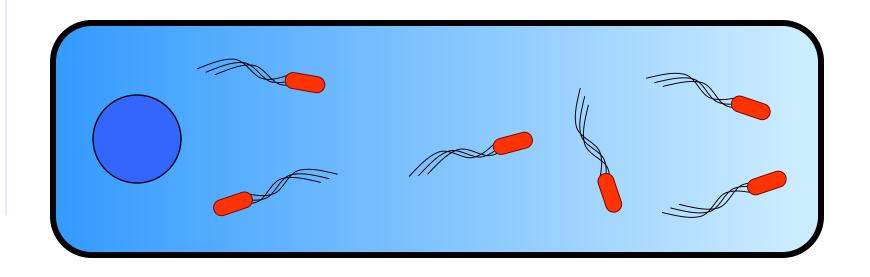
The bacteria communicate their internal oscillator phase with each other using cell-to-cell signaling ...



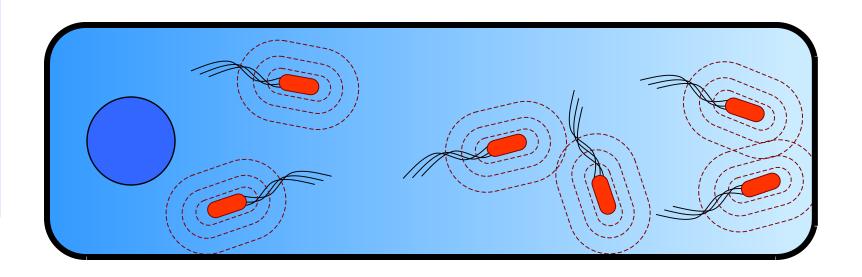
... enabling the entire population to change state synchronously



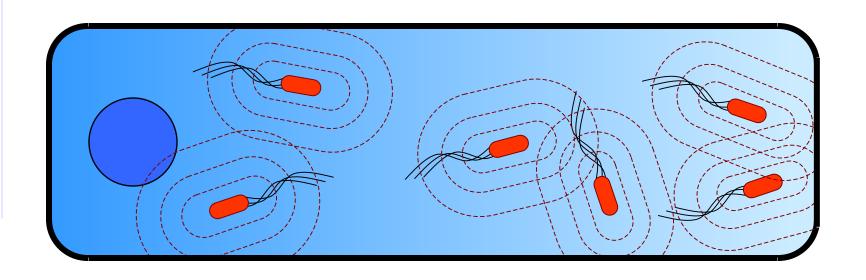
In red bacteria, chemotaxis is disabled
Bacteria start to randomly move away
from the attractant plug



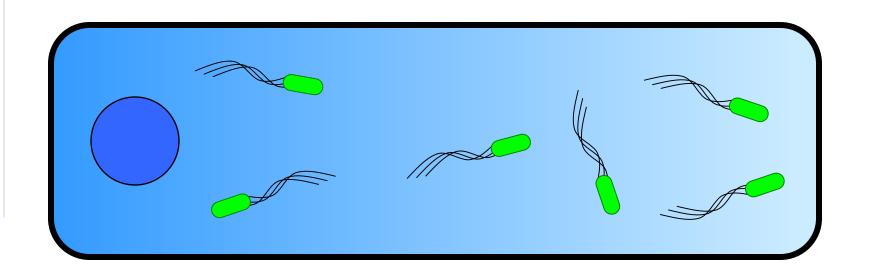
Eventually the bacteria are dispersed around the swimming pool



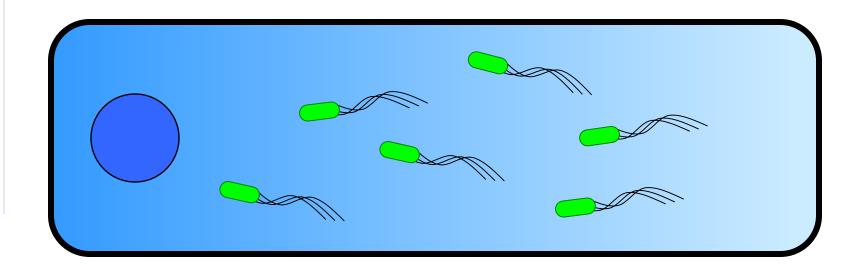
Again the bacteria communicate with each other using cell-to-cell signaling



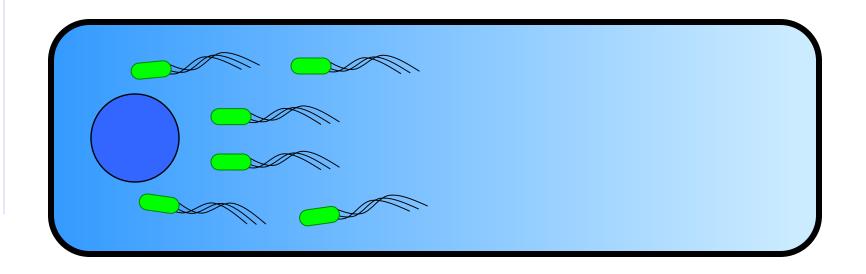
The inter-cellular signaling molecule diffuses throughout out the swimming pool to the entire population



State changes: chemotaxis is enabled

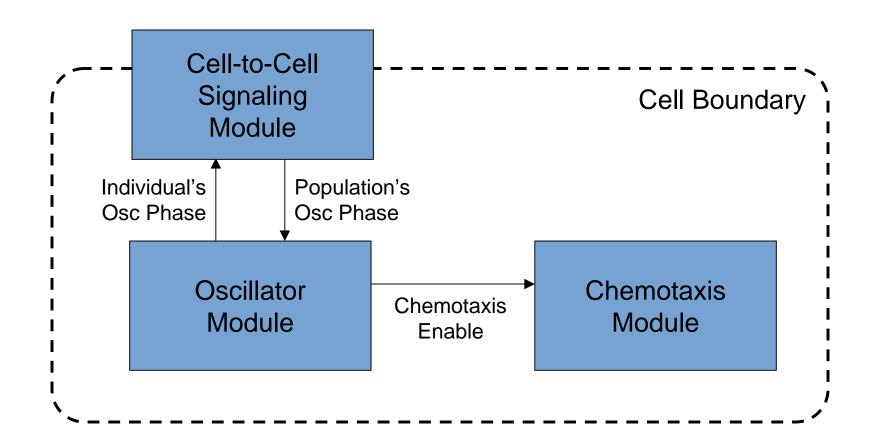


... and onward they swim, doomed to a fate worse than that of Sisyphus

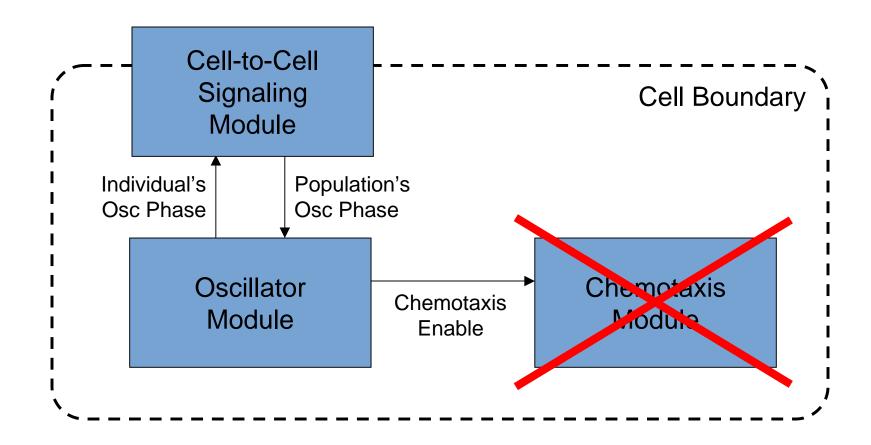


"Futile, this wretched swimming!"

Top-Level System Diagram

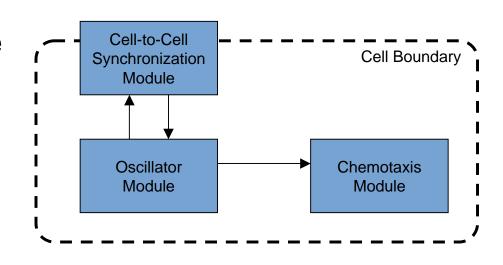


Top-Level System Diagram

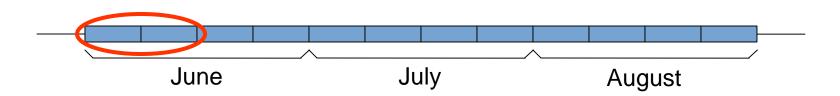


Outline

- Overview
 - Motivation
 - System Description
 - How we got there: Synopsis of Summer Activities
- Module Discussion
 - Cell-to-cell Signaling Module
 - Oscillator Module
 - Chemotaxis Module
 - Module Integration
- Final Remarks



Learning about SynthBio



- Preliminary discussion and design work
- Previous Class Experiences at MIT
 - Much design, little implementation

Cool ideas, but we wondered: "Can we do this?" Had to hit the lab ...

Introduction to a Biology Lab

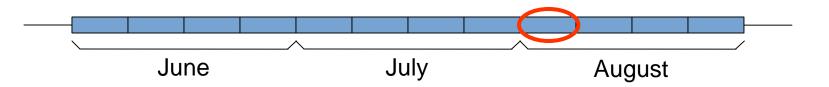


Objective during this period was parts characterization

- Achieved useful work on RBS characterization
- Attempted to build sets of linked inverters
- Began work on cell-to-cell signalling

"So...you're saying that was supposed to be refrigerated?" "Agarose gels, Agar gels...what's the difference?"

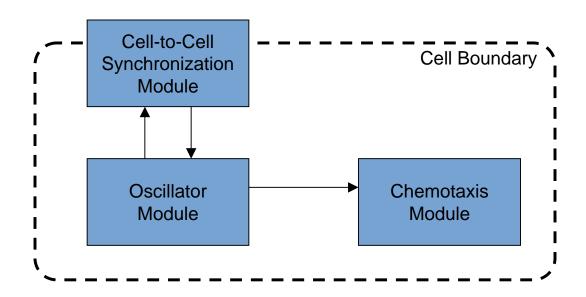
Finalizing Design



Brainstormed several comprehensive and full-system designs

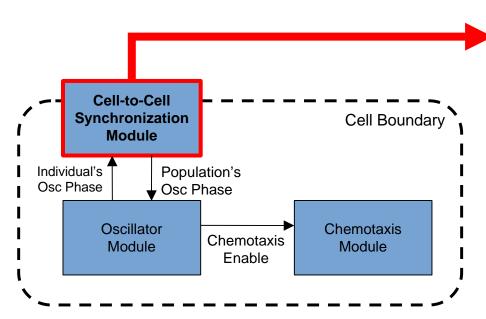
Choosing one design gave us purpose and focus in lab

Thereafter: Making it Happen

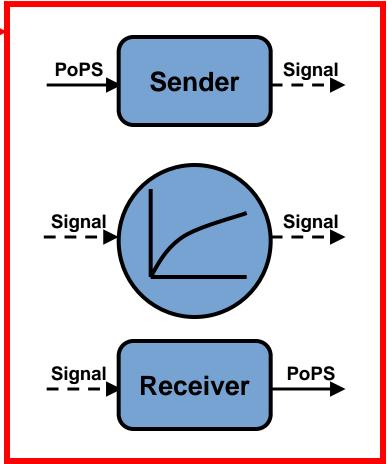


- Final element
 - Integrating modules is surely easier said than done
 - How to prepare the experimental setup for our work?
- Barry, Jason, Fred, and Vikki will now discuss these topics. Chris will offer final remarks.

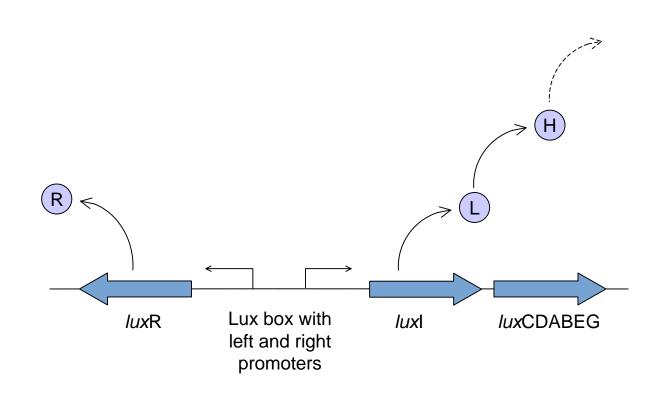
Cell-to-Cell Signaling Objectives



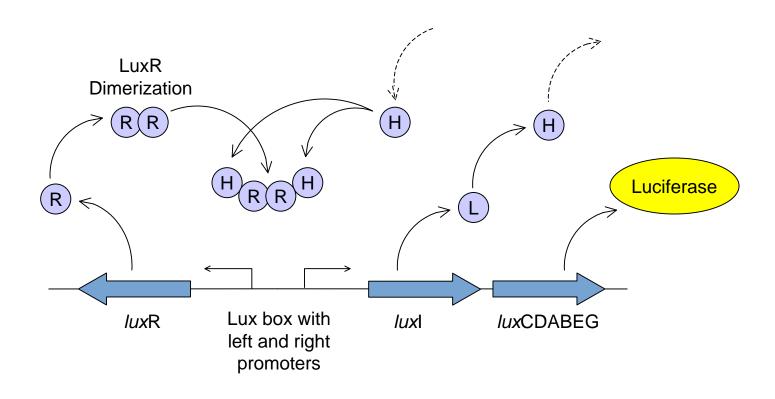
- Decompose signaling into elements
- Design, build and test elements
- Explore how elements might function as part of the full system



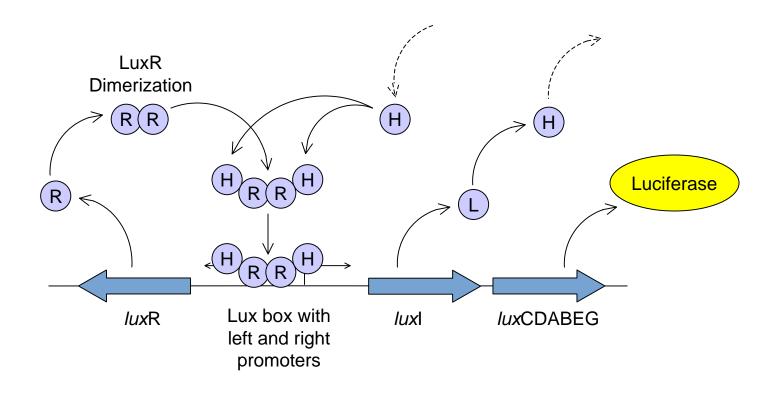
The Lux System



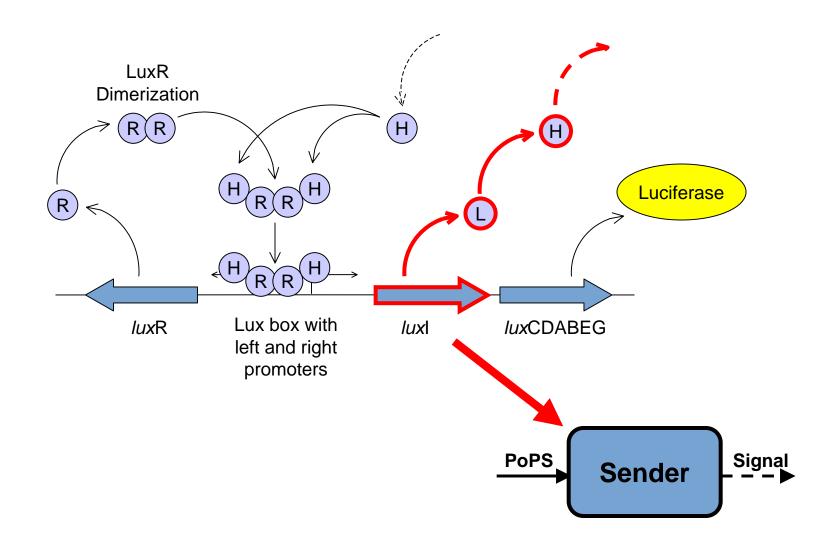
The Lux System



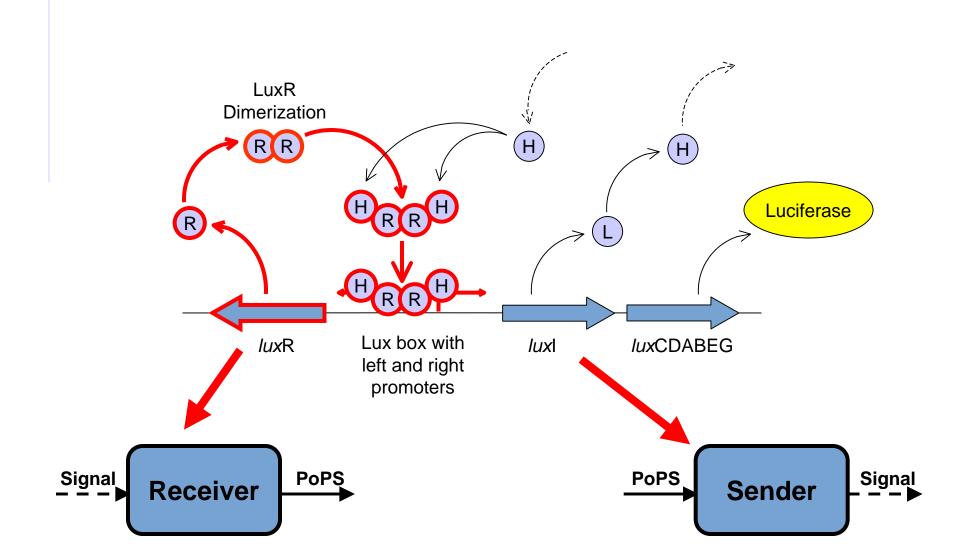
The Lux System



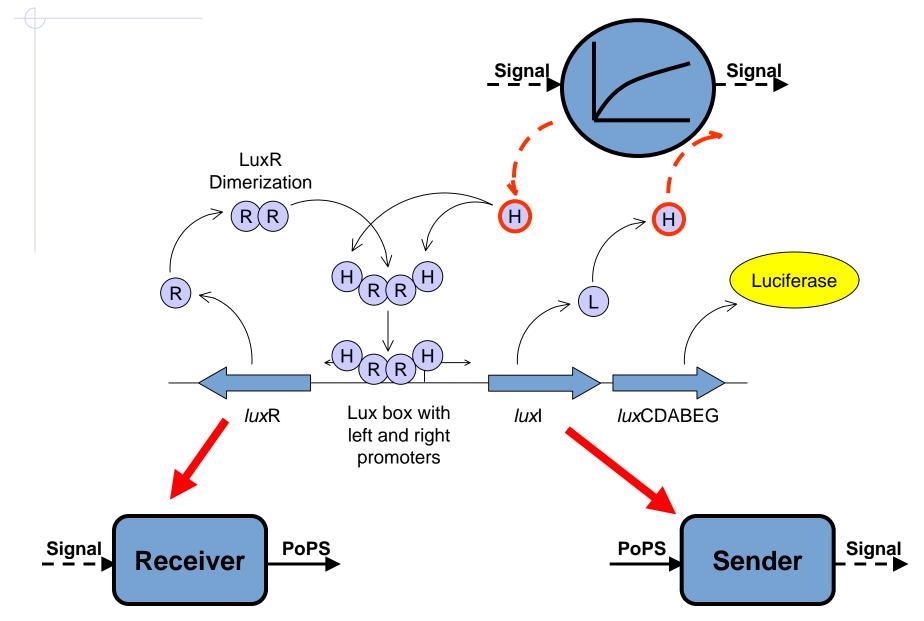
Utilizing Existing Components



Utilizing Existing Components

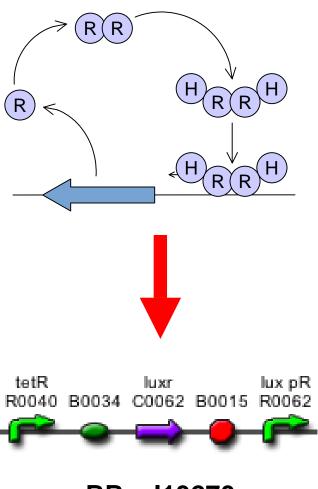


Utilizing Existing Components



Receiver Design

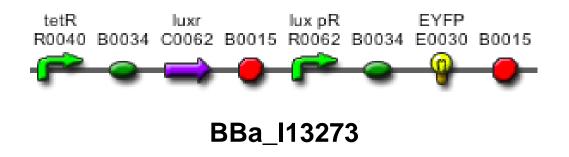




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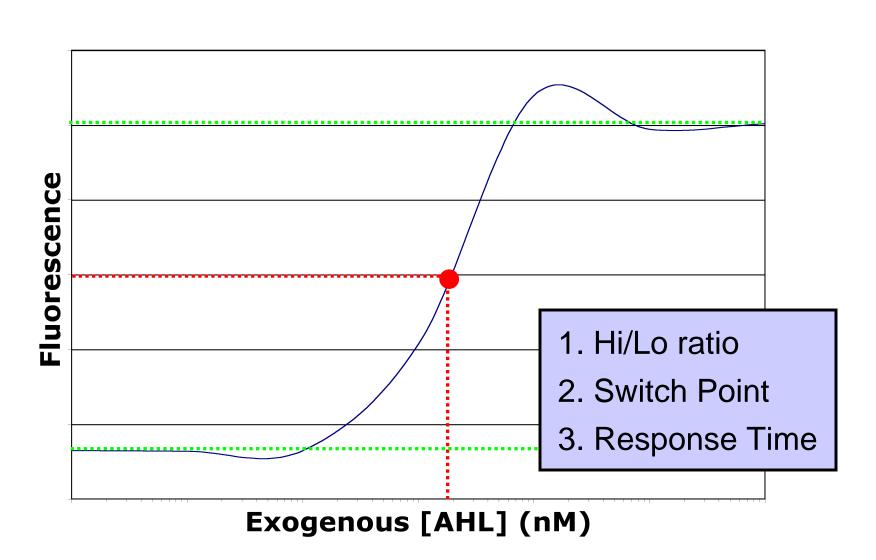
Receiver Building





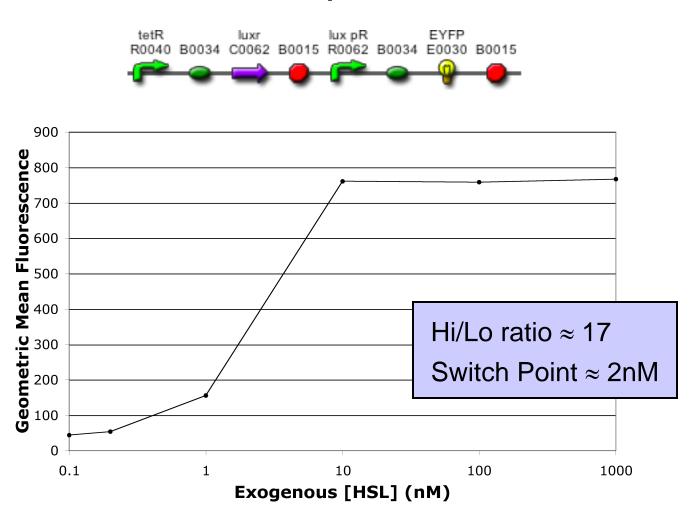
- Varied Upstream Promoter Ptet, luxP_L
- High (100-200) and Low Copy (10-20) Plasmid
- Used YFP Output Device as a PoPS Reporter
- Built in DH5alpha using standardized assembly,
 Transformed into MC4100 and HCB1103





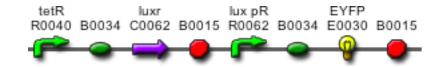


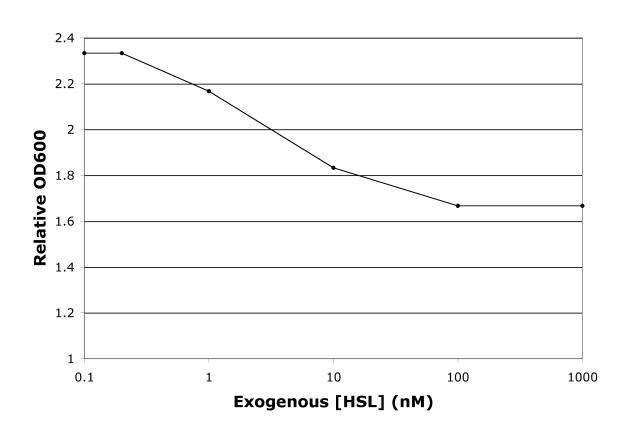
113273 - pSB1A2





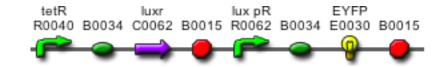
113273 - pSB1A2 - Growth Defects

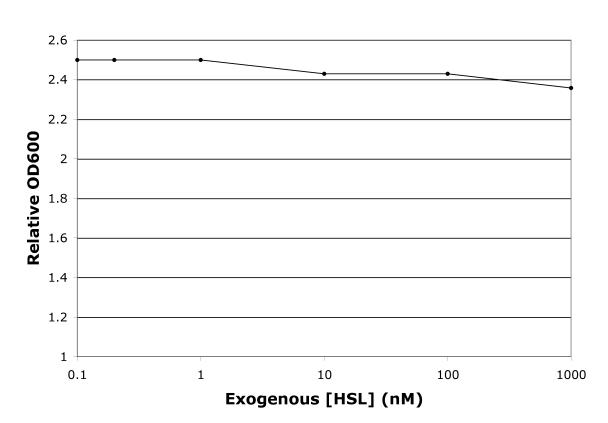






113273 - pSB3K3 - Growth Restored

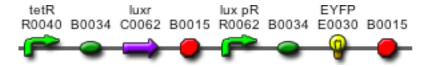


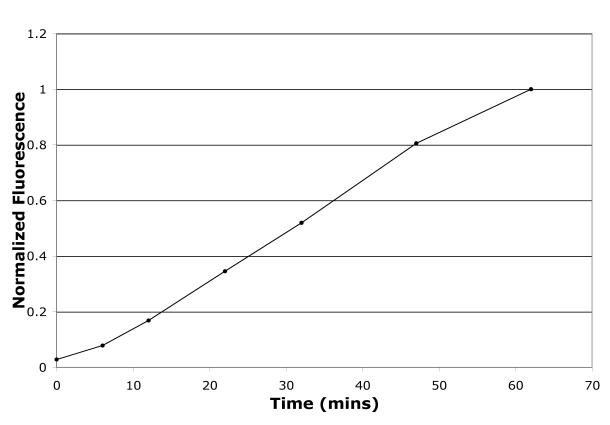


Receiver Testing



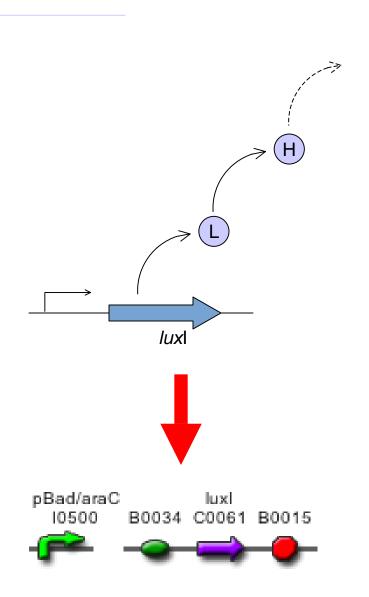
113273 - pSB3K3 - Response Time

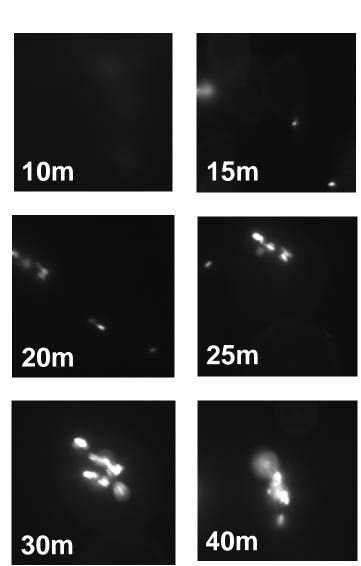




Senders





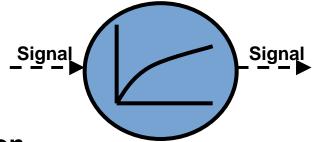


Transmission

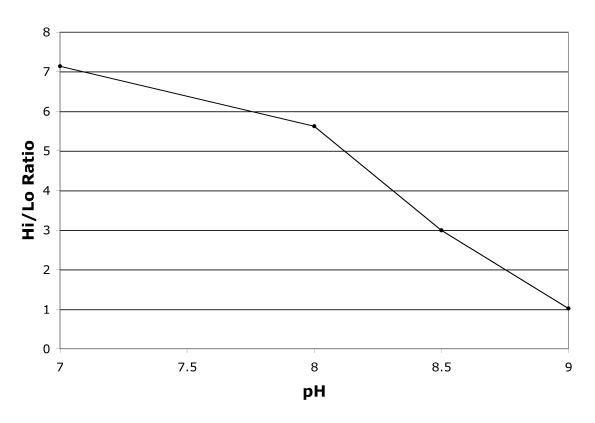
Signal

- Diffusion rate
- Degradation due to dilution (e.g. in chemostat)
- Degradation due to raised pH
- Active enzymatic degradation aiiA

Transmission

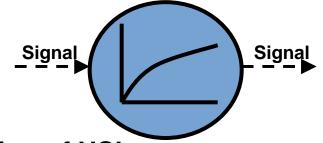


pH Dependent Degradation



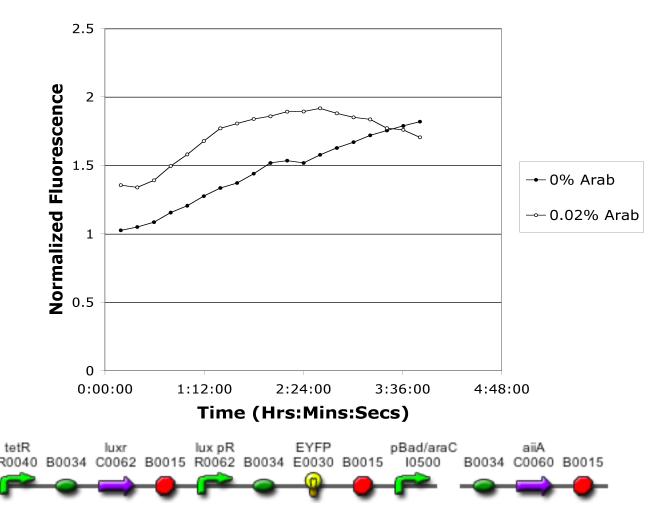
60 minute incubation of HSL at various pH Use that HSL to activate receivers at neutral pH

Transmission



aiiA

Enzymatic intracellular Degradation of HSL

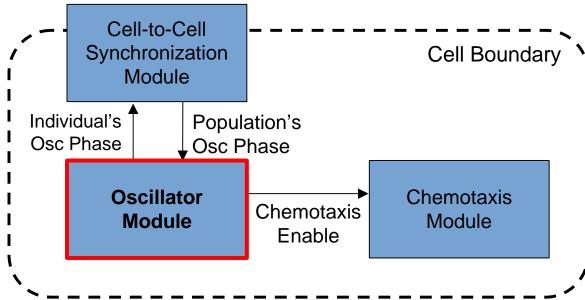


Future Work

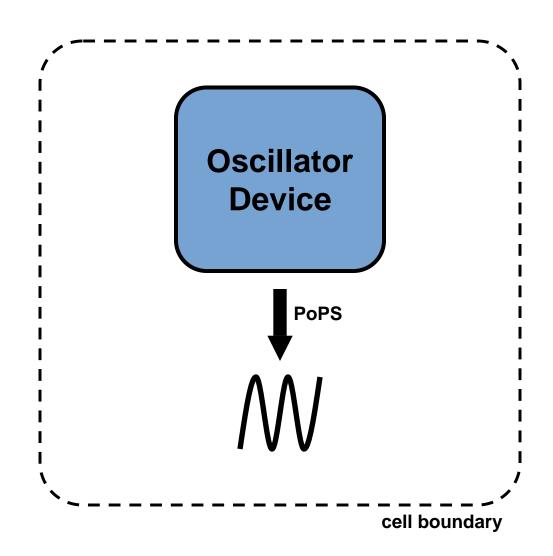
- Develop the ability to adjust receiver transfer function parameters at will
- Complete characterization of existing sender device using the receiver device
- Build and test the sender device used in the synchronized oscillator
- Continue to test the aiiA degradation mechanism
- Quantify parameter robustness under different operating conditions – chemostats, microscopes etc.

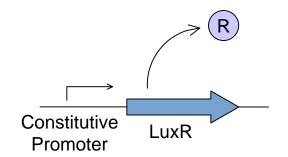
Oscillator Module

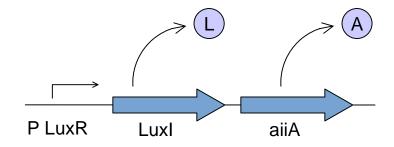
- Stand-alone Oscillator
 - Relaxation Oscillator
 - Ring Oscillator
- Synchronized Oscillator
 - Synchronators
 - Synchronized Ring Oscillator
- Future Work



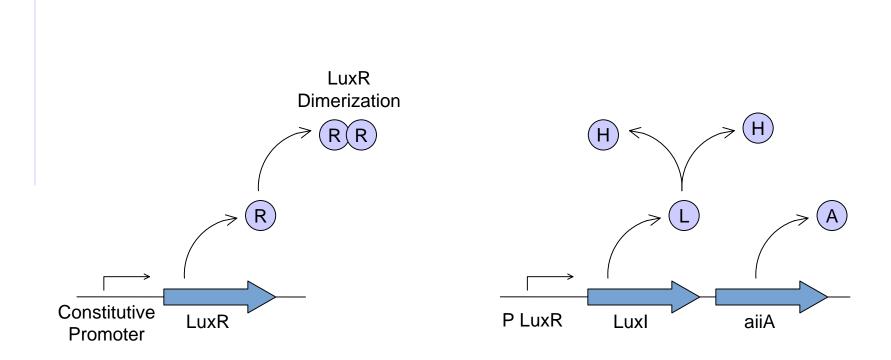
Input/Output



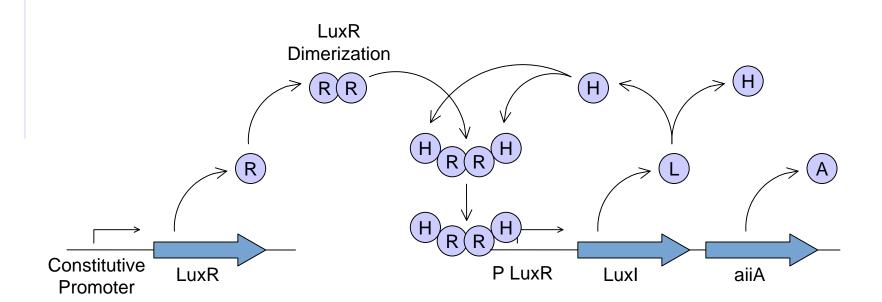




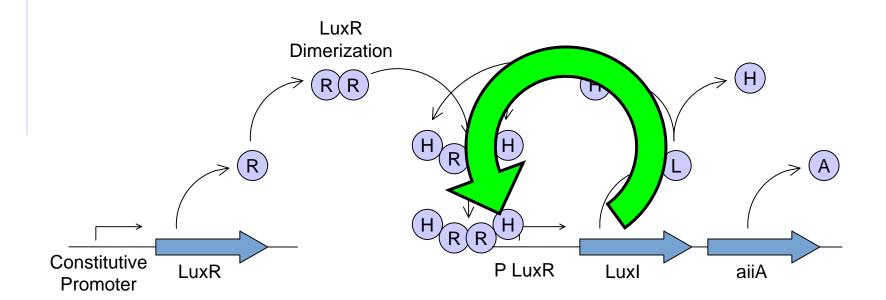
LuxR is constitutively expressed, while LuxI and aiiA are regulated by a LuxR activated promoter



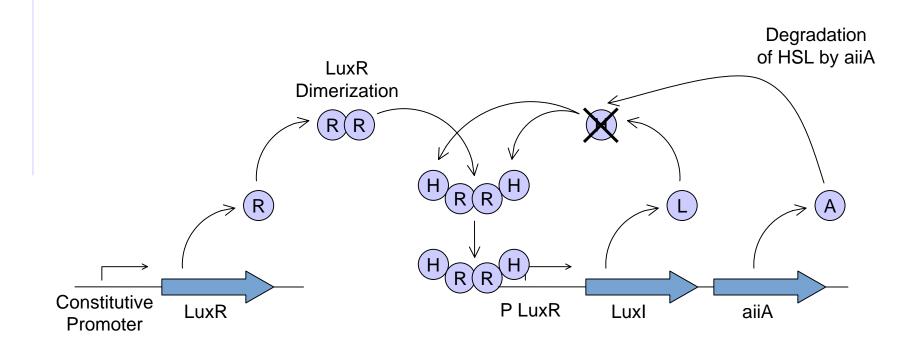
LuxR forms a dimer while LuxI synthesizes HSL



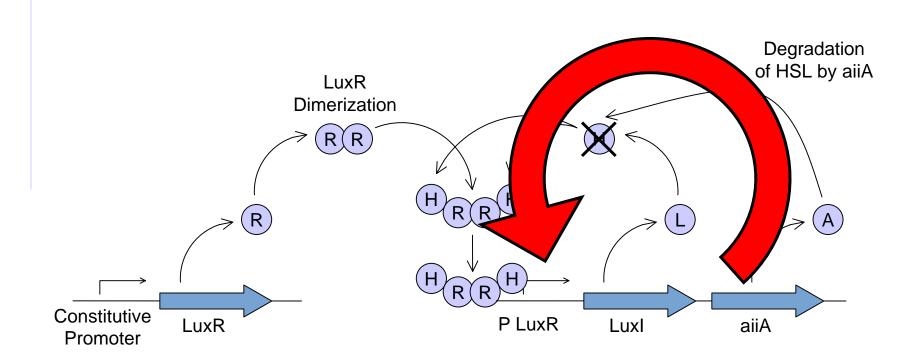
LuxR and HSL bind to form the transcriptional activator providing positive feedback



LuxR and HSL bind to form the transcriptional activator providing positive feedback



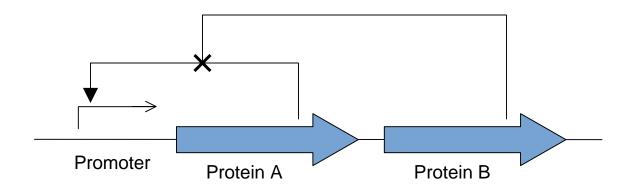
aiiA degrades HSL providing negative feedback



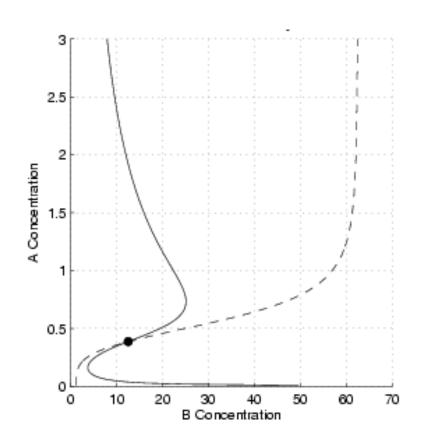
aiiA degrades HSL providing negative feedback

Simplified Relaxation Oscillator

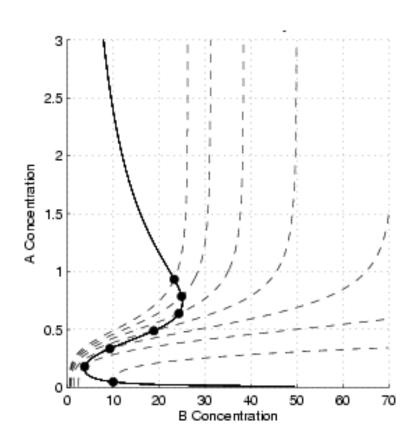
- Initial modeling work used a system of continuous differential equations to examine a simplified oscillator
- Folds the positive feedback into a single Protein A ignoring the details of LuxI, HSL, and LuxR
- Even with these simplifications, the model can give insight into what experimental constructs would be useful when building the actual Lux/aiiA oscillator



State Space Analysis

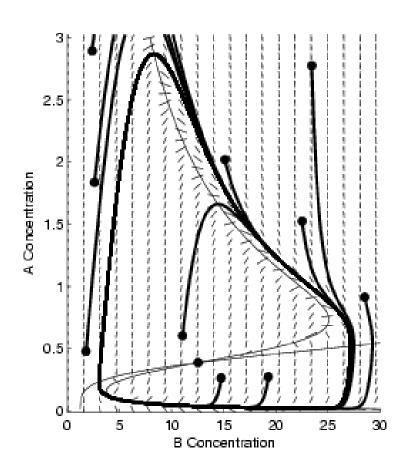


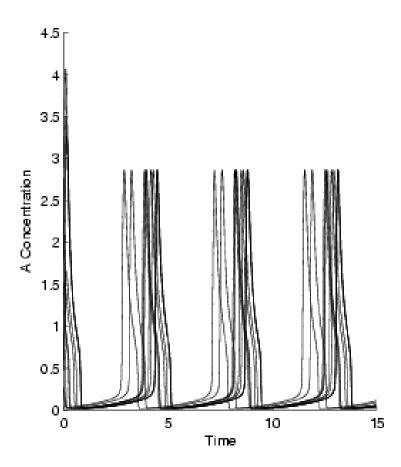
Intersection of nullclines yields system equilibrium point



Equlibrium point changes with Protein B degradation rate

Preliminary Modeling Results





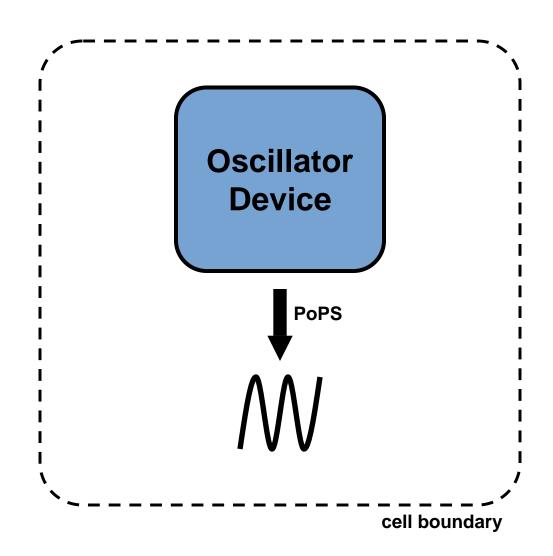
A vs B State Space

Concentration of A vs Time

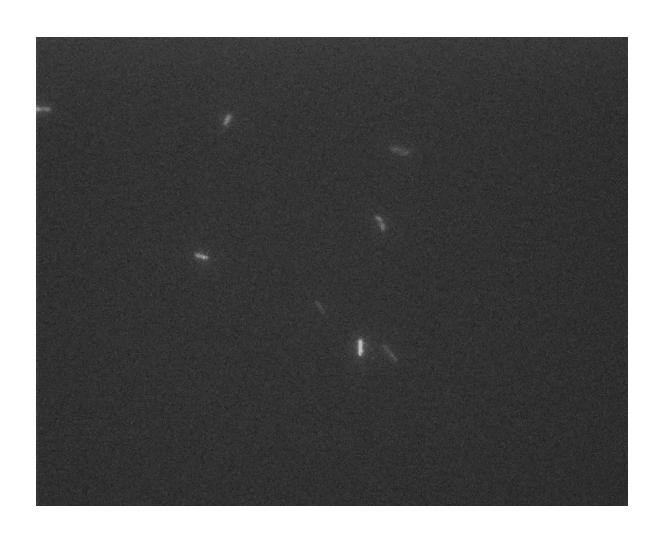
Experimental Work

- Modeling work suggested possible test constructs
- Experimental work on the Lux/aiiA relaxation oscillator was put on hold
 - Initial results on aiiA were discouraging
 - Not enough degradation tags were available to effectively tune the aiiA degradation rate

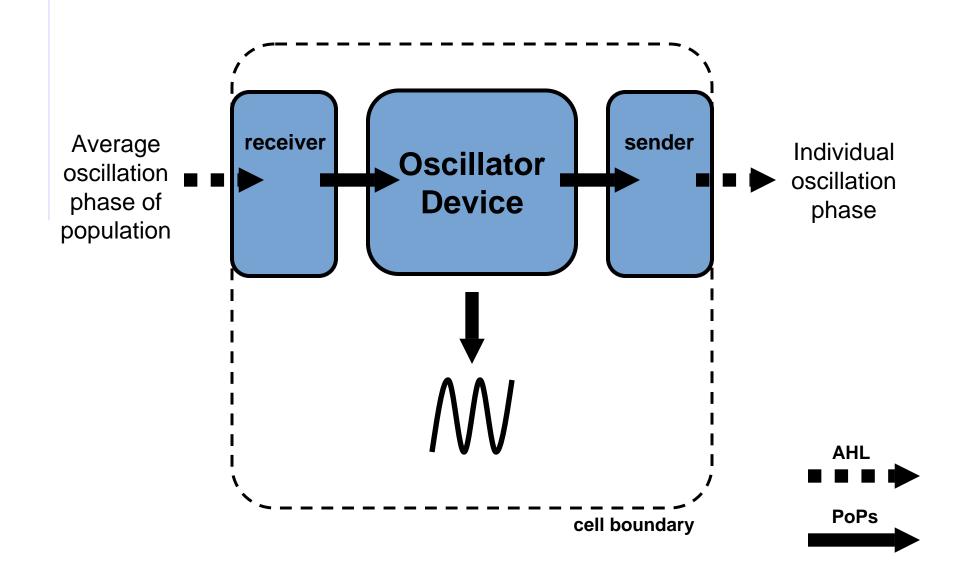
Input/Output



Ring Oscillator



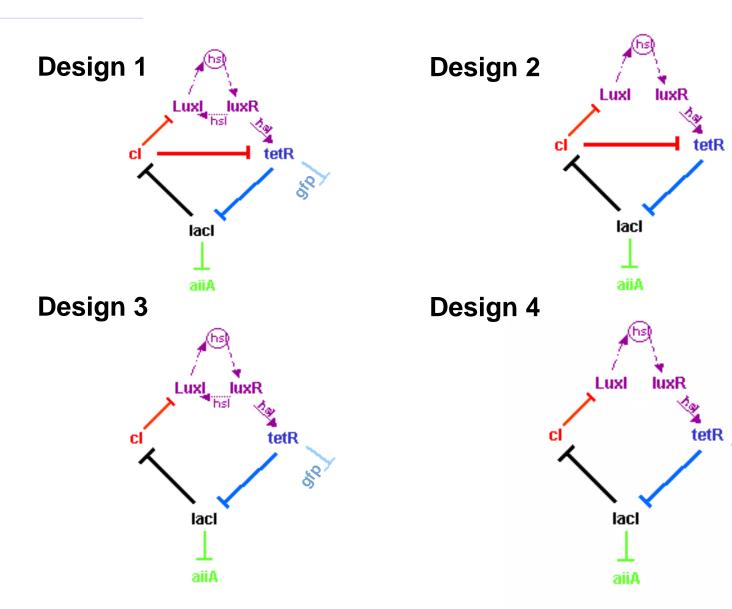
Input/Output



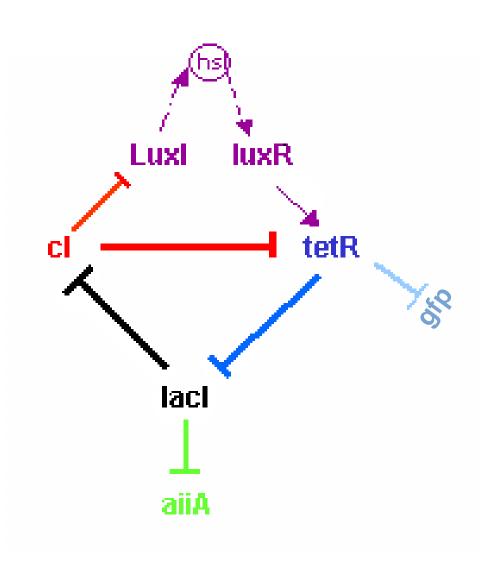
Synchronized Oscillator Options

- Repressilator & Synchronization Device
 - Functional oscillator
 - Need to design synchronization device
- Synchronator
 - 4 designs available from the MIT 2003 Synthetic Biology course
 - Designed to synchronize, completely built, but untested and uncharacterized
- See-ya-lator
 - Modeled after Yankees playoff performance

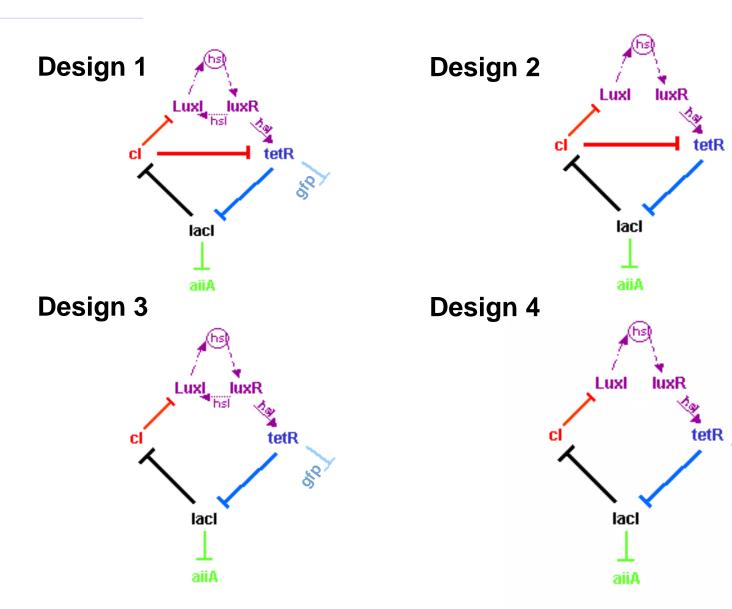
Synchronator Designs



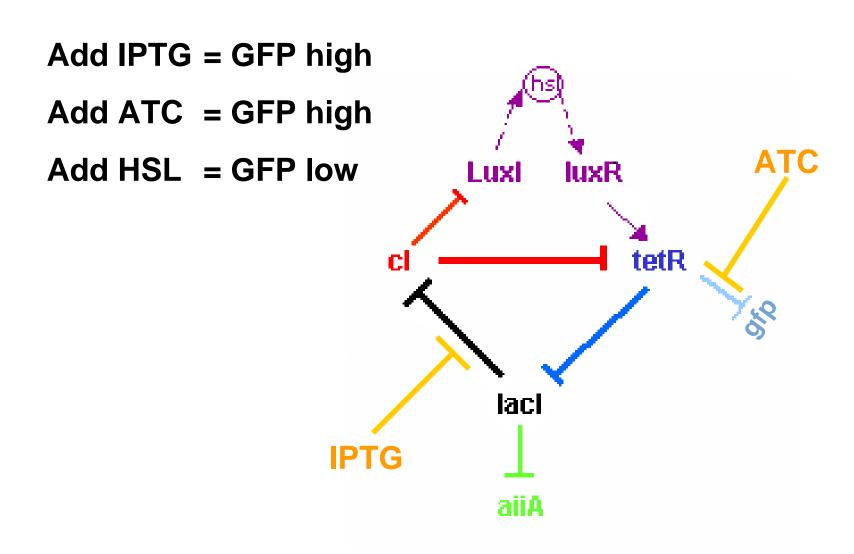
Synchronator Design 2



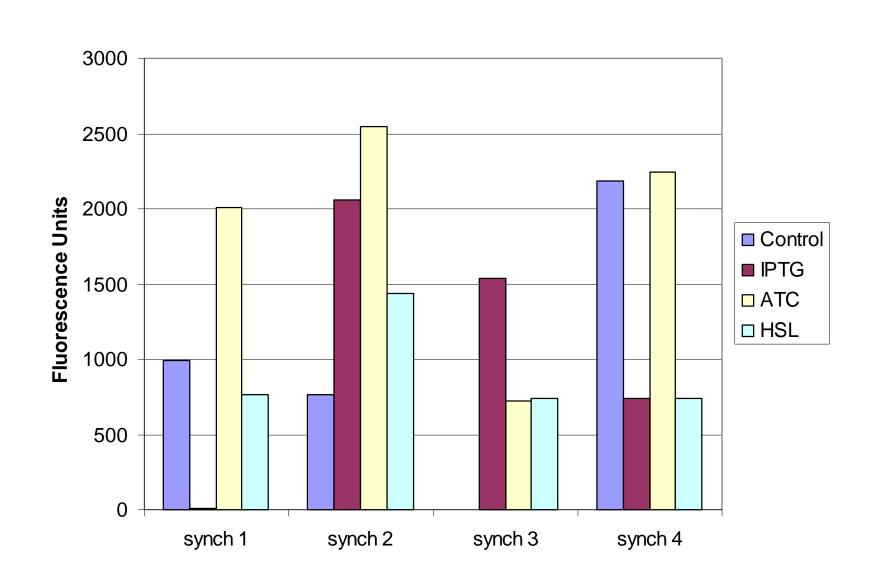
Synchronator Designs



Oscillator Lockdown Experiment



Synchronator Lock-Down

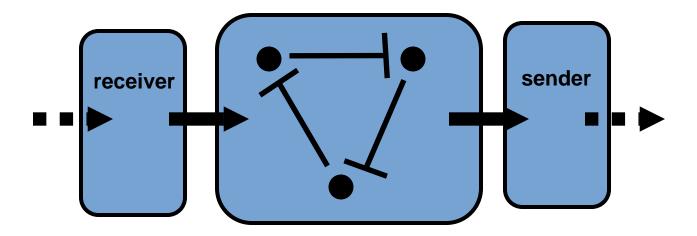


Synchronator 2 Movie



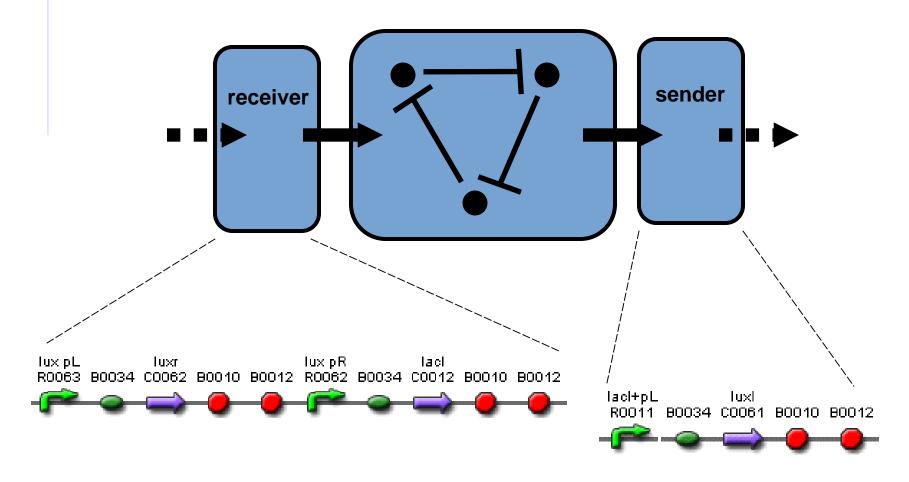
Synchronized Ring Oscillator

 Add a synchronization element to the Repressilator (Garcia-Ojalvo, Elowitz, Strogatz, PNAS 2004)

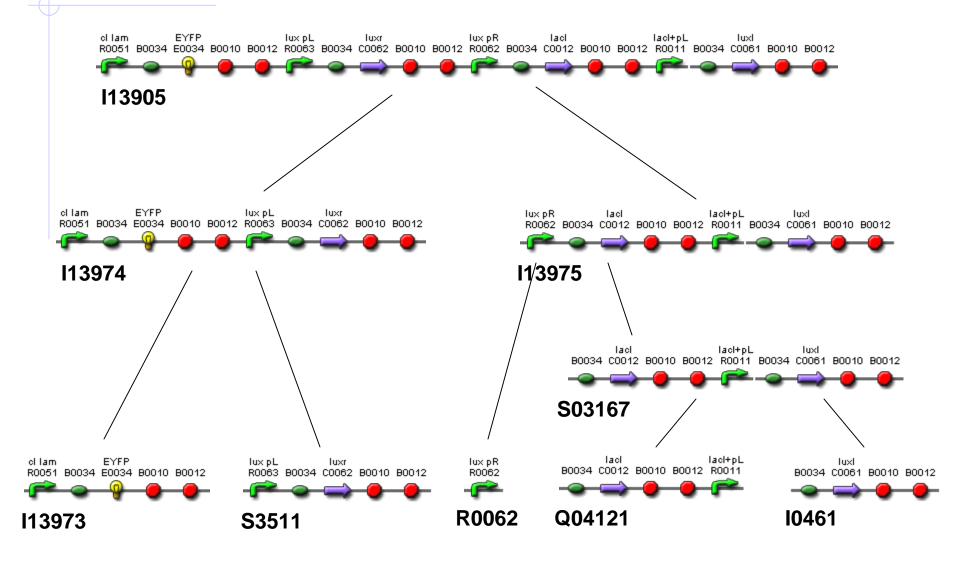


Synchronized Ring Oscillator

 Add a synchronization element to the Repressilator (Garcia-Ojalvo, Elowitz, Strogatz, PNAS 2004)



Construction of Synchronization Device

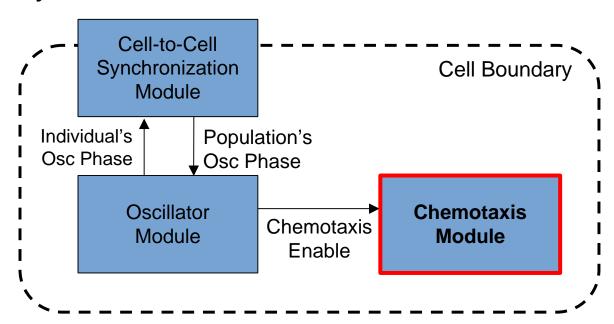


Future Work

- Synchronized Ring Oscillator
 - Lock-down experiments
 - Agarose Pad Time Lapse Movie
 - Continuous Culture (chemostat)
 - Plate Reader Time Course
- Relaxation Oscillator
 - Explore aiiA further to determine why it isn't functioning as expected
 - Build test constructs and characterize

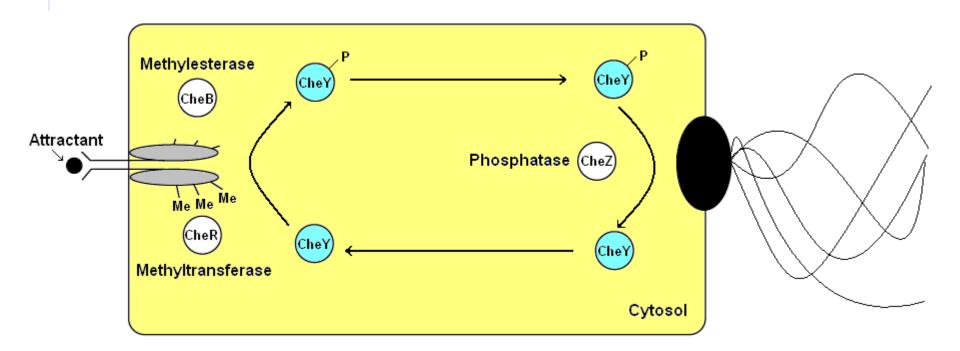
Chemotaxis Module

- Chemotaxis biology
- Chemotaxis devices
 - Restoring motility
 - Deactivating motility
- Chemotaxis assay
- Results
- Future work



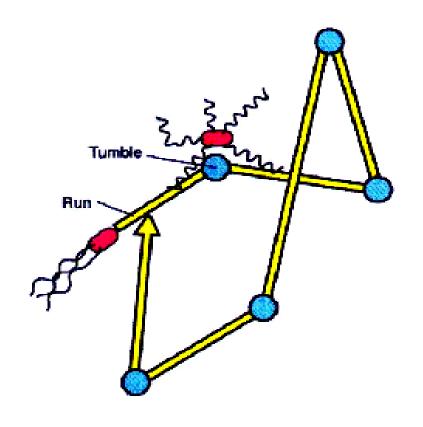
Chemotaxis in Escherichia Coli

- Four intracellular signaling proteins
 - CheB, CheR, CheY, and CheZ
 - Maintained at specific levels and ratios



Motile Behavior

- Net movement toward or away from chemicals result from the combined effect of smooth runs and tumbles
- The expression and activity of signaling proteins (CheB, CheR, CheY and CheZ) determines the frequency of tumbles and runs



Too Much or Too Little?

Absence of any signaling protein affects motile behavior

Genotype	Motility	Phenotype
wt	+	smooth runs and tumbles
∆CheB	+	tumbles
∆CheR	+	smooth runs
∆CheY	-	none
∆CheZ	+	smooth runs

Overexpression of any signaling molecule affects motile behavior

Genotype	Overexpression	Phenotype
wt	CheB	smooth runs
wt	CheR	tumbles
wt	CheY	tumbles
wt	CheZ	smooth runs

Too Much or Too Little?

Absence of any signaling protein affects motile behavior

Genotype	Motility	Phenotype		
wt	+	smooth runs and tumbles		
∆CheB	+	tumbles		
∆CheR	+	smooth runs		
∆CheY	-	none		
∆CheZ	+	smooth runs		

Overexpression of any signaling molecule affects motile behavior

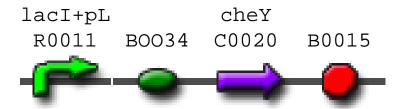
Genotype	Overexpression	Phenotype
wt	CheB	smooth runs
wt	CheR	tumbles
wt	CheY	tumbles
wt	CheZ	smooth runs

CheY concentration in RP437: $8,200 \pm 310$ per cell in rich media and $6,300 \pm 70$ per cell in minimal media

Li M, Hazelbauer G "Cellular Stoichiometery of the Components of the Chemotaxis Signaling Complex" Journal of Bacteriology, 2004, 186(12) 3687-3694

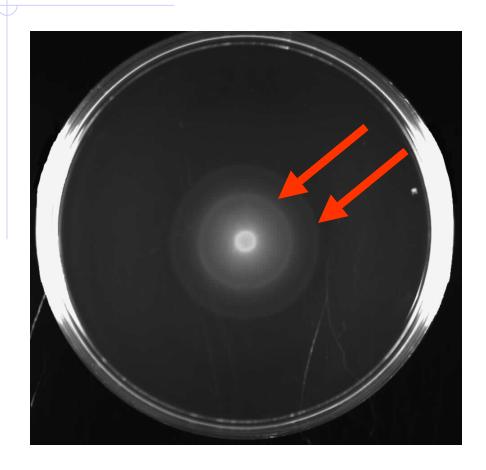
Building Chemotaxis Output

CheY quadparts: High Copy (150-200) and Low Copy (15-20)



- Two methods for coupling to Chemotaxis
 - Restoration of normal chemotaxis in ∆CheY mutant strain
 - Deactivation of normal chemotaxis in wild type strain
- Characterizing quadpart expression
 - Time frame of protein expression
- Observing the inactivation and deactivation of bacterial chemotaxis
 - Swarm plate characterization
 - Drop assay and bacterial clustering on glass slide
 - Capillary Assay

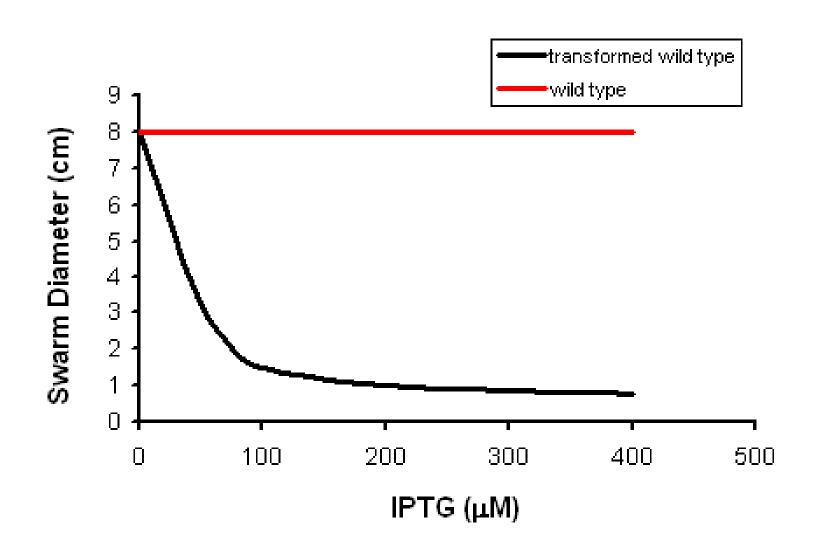
Swarm Plate Assay



10ul of bacterial suspension 30°C for duration of swarming

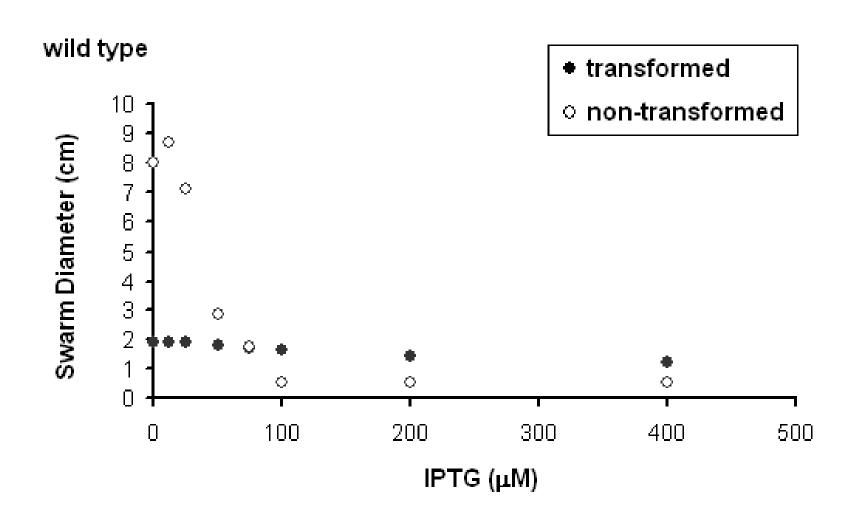
- Amino acids are bacterial chemoattractants
- Nutrient consumption produces gradient
- Ring formation on agar corresponding to particular amino acid/chemoattractant consumed by motile bacteria
- Movement away from the center (point of inoculation)

Deactivation: Expected Results



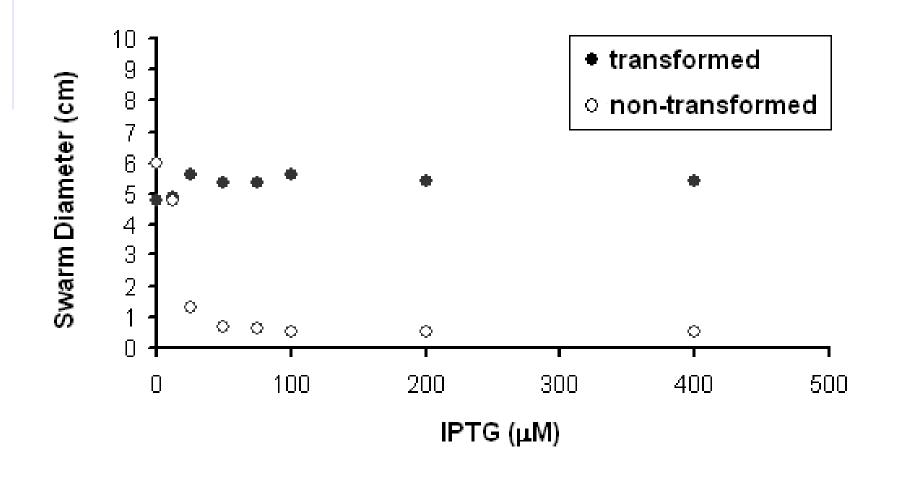
Deactivation: High Copy Results

High copy CheY expression in wild type



Deactivation: High Copy Results

High copy CheY expression in LacI- strain (wild type background)



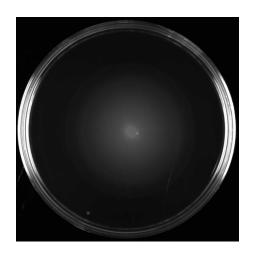
Deactivation: Low Copy Results

25μM IPTG induction for 2 hours, $OD_{660} = 0.1$ 10ul spot, **16 hrs**

Lacl⁻

Expected: Swarm

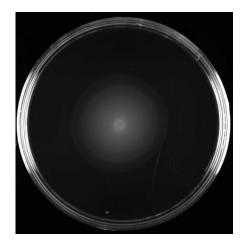
Observed: Swarm



wild type

Expected: Swarm

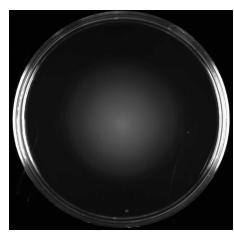
Observed: Swarm



Lacl⁻ (transformed)

Expected: Swarm/No rings

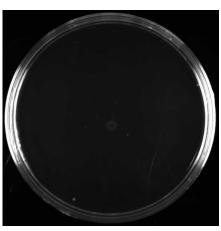
Observed: Swarm/No rings



wild type (transformed)

Expected: Swarm

Observed: No Swarm

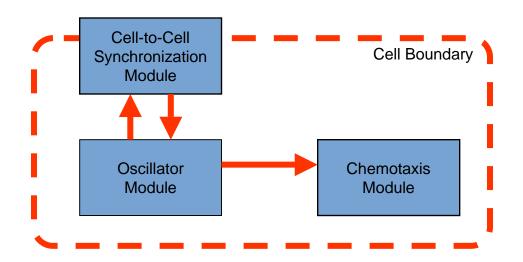


Future Work

- More low copy construct (15-25 per cell) experiments
- Experimentation with ∆cheY mutants of E. coli RP437 for the restoration of motility
 - Requires the tuned expression of CheY
- Drop assay or coverslip assay to observe bacterial aggregation
 - Characterization of immediate chemotaxis response under varying levels of induction
- Coupling to population oscillator module and cell-cell signaling module

Module Integration

- Operating Conditions
 - Strain
 - pH
 - Temperature
 - Test setup
- Inter-Module
 Communication
 - Signal Interpretation
 - Timing



Op Conditions: Strain

Module	Known Requirements	Tested Conditions		
Cell-Cell Signaling		MC4100, DH5alpha		
Oscillator	Lacl-	MC4100		
Chemotaxis	Chemotactic	RP437 (HCB33)		

- Testing Cell-Cell signaling module in RP437
- Create Lacl- version of RP437
- Test combined construct in Lacl- version of RP437

Op Conditions: pH and Temp

Component	Known Requirements	Tested Conditions			
Cell Cell Signaling	HSL stability is pH dependent	37° Celsius, pH 7			
Oscillator		37° Celsius, pH 7			
Chemotaxis	pH around 7, temperature around 30° Celsius	30° Celsius, pH 6.5-7.5			

Future Plans

Test combined system in pH7, 32° Celsius

Op Conditions: Test Setup

Component	Known Requirements	Tested conditions
Cell Cell Signaling		Culture tubes
Oscillator		Chemostat Agarose pads
Chemotaxis	Steady gradient of chemoattractants	Swarm plates

- Plan I: Swimming Pool for continuous observation
- Plan II: Time Course Sampling with time course test for chemotaxis

Inter-Module: Signal Interpretation

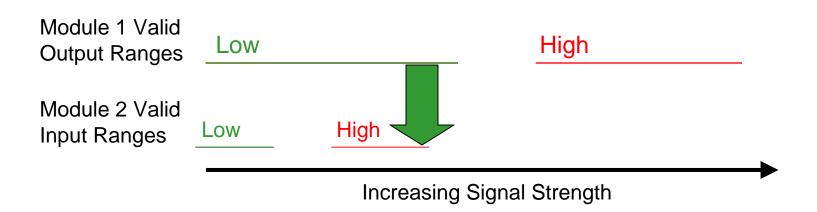
Signals given as a logical "1" or "0" output from one module must be interpreted as a logical "1" or "0' input by the other modules

Module 1 Valid Output Ranges	Low	High	
Module 2 Valid Input Ranges	Low	High	
		Increasing Signal Strength	

- Do signal strength characterization tests for modules
- If strengths don't match, fine tune by swapping promoter/RBS

Inter-Module: Signal Interpretation

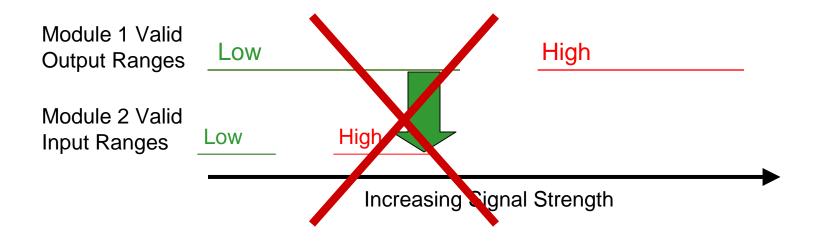
Signals given as a logical "1" or "0" output from one module must be interpreted as a logical "1" or "0' input by the other modules



- Do signal strength characterization tests for modules
- If strengths don't match, fine tune by swapping promoter/RBS

Inter-Module: Signal Interpretation

Signals given as a logical "1" or "0" output from one module must be interpreted as a logical "1" or "0' input by the other modules



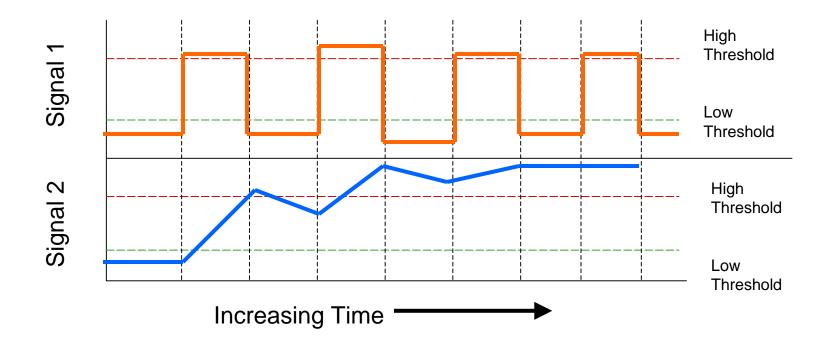
- Do signal strength characterization tests for modules
- If strengths don't match, fine tune by swapping promoter/RBS

Inter-Module: Timing

Timing of oscillator signals must match up with timing of cellcell signaling and chemotaxis modules

Signal 1: (Oscillator) Rise Time=0; Fall time=0; Period =2

Signal 2: (Follower) Rise Time=1; Fall time=2



Inter-Module: Timing

 Cell-to-Cell Signaling and Chemotaxis must be able to turn on and off in at most the amount of time it takes the Oscillator to turn on and off.

Current Knowledge

- Repressilator : Period ~2 hours
- Cell-to-Cell and Chemotaxis rising edge is fast
- Cell-to-Cell and Chemotaxis falling edge is slow

- Try getting the HSL signals to degrade faster by operating at a higher pH (10) or in a chemostat
- Characterize off times for chemotaxis module

Integration Summary

- Integration is HARD!
 - Operating Conditions
 - Inter-module Communication
- Still a lot of work to be done

Final Remarks

- Overview of summer accomplishments
- Advice for future summer competitions
- Key enablers for the field of synthetic biology
 - Assembly process
 - Device characterization
 - Standard operating conditions

Summer Accomplishments

- Over 200 new BioBrick parts added to the registry
- Device characterization
 - RBS measurements
 - Preliminary copy number measurements
 - Basic chemostat constructed and tested
 - Many inverter measurement constructs ready to be tested
- Cell-to-cell signaling module
 - Working Lux sender/receiver constructed with BioBrick parts
 - Characterizing receiver transfer curves
 - Verified importance of low-copy constructs
 - Characterization of cell-to-cell signaling channel

Summer Accomplishments

Oscillator module

- Modeling work on Lux/aiiA relaxation oscillator
- Refined techniques for creating time-lapse movies
- Verified repressilator ring oscillator
- Tested previously designed Synchronators
- New synchronized repressilator is built and ready for testing

Chemotaxis module

- Working swarm plate chemotaxis assay
- Results on possibility of transcriptional control of chemotaxis
- Twelve synthetic biology students who are excited about the potential of this new field
- Five extremely frustrated advisors who are looking forward to a long winter vacation

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C0024	I13015	I13264	I13314	I13019	I13107	I13637	I13210	E0669	Q00121
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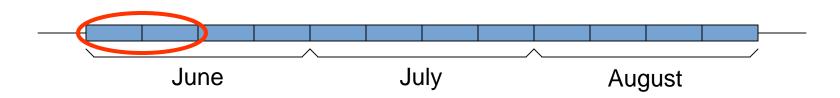
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	I13005	I13213	I13309	C0261	I13102	<u> 113626</u>	I13013	I13974	I13851	
	I0466	I13220	I13310	10463	I13103	I13627	I13006	I13975	I13971	
	I0467	I13261	I13311	10464	I13104	I13631	C0056	I13976	I13972	
ĺ	10468	I13262	I13312	I0465	I13105	I13633	I13016	I13977	I13973	

C0020	I13000	I13263	I13313	I13014	I13106	I13634	I13209	E0130	P0474
C0024	I13015	I13264	I13314	I13019	I13107	I13637	I13210	E0669	Q00121
C0028	I13018	I13265	I13910	I13020	I13108	I13644	I13900	I13600	Q00400
I13700	I13034	I13266	I13911	I13021	I13109	I13645	I13901	I13601	Q02121
I13701	I13035	I13270	I13912	I13022	I13110	I13647	I13902	I13602	Q02400
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G00701	I13205	I13304	I534160	I13031	I13617	I13008	I13941	I13062	
I13001	I13206	I13305	I13207	I13032	I13621	I13009	I13942	I13072	
I13002	I13208	I13306	C0063	I13033	I13623	I13010	I13943	I13800	
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I13005	I13213	I13309	C0261	I13102	I13626	I13013	I13974	I13851	
10466	I13220	I13310	10463	I13103	I13627	I13006	I13975	I13971	
10467	I13261	I13311	I0464	I13104	I13631	C0056	I13976	I13972	
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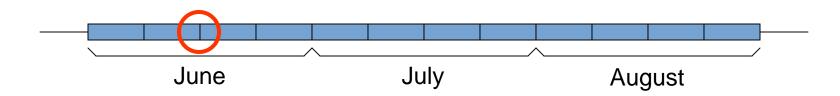
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C0028	I13018	I13265	I13910	I13020	I13108	I13644	I13900	I13600	Q00400
I13700	I13034	I13266	I13911	I13021	I13109	I13645	I13901	I13601	Q02121
I13701	I13035	I13270	I13912	I13022	I13110	I13647	I13902	I13602	Q02400
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I13005	I13213	I13309	C0261	I13102	<u>113626</u>	I13013	I13974	I13851		
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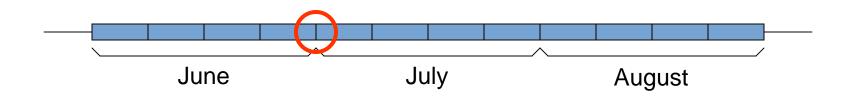
Structured introductory two-week curriculum

- Daily lectures in the mornings and specific lab tutorials in the afternoons
- Students would model, assemble, and characterize a simple synthetic system such as a single OR gate
- Teaches synthetic biology basics and experimental lab technique as well as providing a solid foundation for initial design work



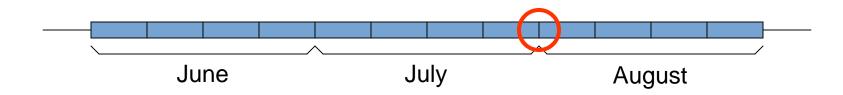
More milestones and incremental deliverables

Report on simple synthetic system



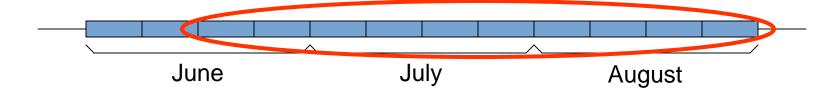
More milestones and incremental deliverables

- Report on simple synthetic system
- Preliminary design specification



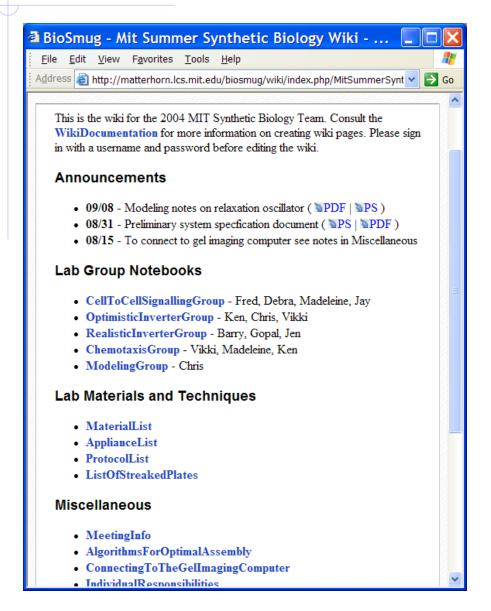
More milestones and incremental deliverables

- Report on simple synthetic system
- Preliminary design specification
- Interim progress report



More milestones and incremental deliverables

- Report on simple synthetic system
- Preliminary design specification
- Interim progress report
- Periodic logs kept by each student and lab-group

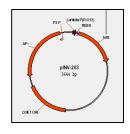


Inter-team collaboration

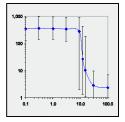
- Periodic conference calls
- Distribute design specifications and interim reports to all teams
- Logs managed in online forum accessible by all teams

Key Enablers for SynthBio

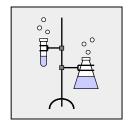
Through our summer experiences we identified three key enablers which will greatly help future work in the field of synthetic biology



Assembly Process

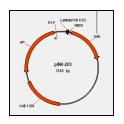


Device Characterization



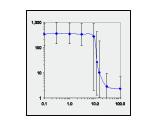
Standard Operating Conditions

Assembly Process



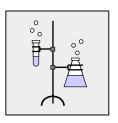
- Remarkable that a group of students with very little biology background was able to build working biological parts relatively quickly
- Even so, current assembly process placed significant constraints on what was possible
 - Took on average one week per stage
 - When assembly failed very difficult to determine why
- Assembly is an important research topic
 - Optimize each stage
 - Characterize and model error rates
 - Develop more assembly tools

Device Characterization



- Modeling work is significantly hampered by lack of useful device characterization
- Device characterization is challenging
 - What do we actually measure?
 - How do we measure it?
 - How do we make measurements repeatable?
- Accurate device characterization will enable
 - Effective parameterized models
 - More rational design
 - Easier reuse of previously developed parts

Operating Conditions



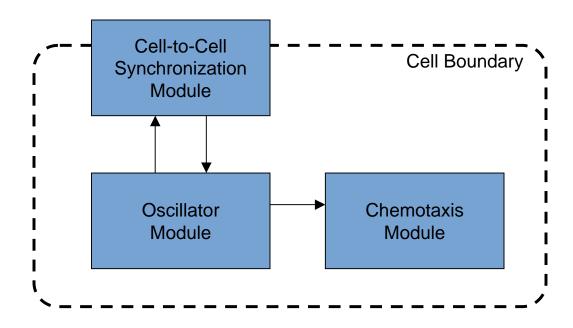
- Currently no standard operating conditions
 - Strain, media, growth phase are usually documented but they still vary with each experiment
 - Standard conditions enable easier result comparisons
 - Standard operation conditions also make it easier to predict how future systems will behave
- Standard operating conditions is challenging
 - Difficult to choose a single set of conditions since different experiments have different requirements
 - Continuous culture using chemostat is an attractive possibility but needs more work

Acknowledgements

- Strains + Plasmids
 - Howard Berg (Harvard)
 - Karen Fahrner (Harvard)
 - Michael Elowitz (CalTech)
- Cell-to-Cell Signaling Advice
 - Ron Weiss (Princeton)
- Assembly
 - Caitlin Conboy
 - Jen Braff
- Advisors
 - Drew Endy
 - Randy "Registry Ranger" Rettberg
 - Gerry Sussman
 - Pam Silver
 - Tom Knight

Conclusions

- We have designed and made strong progress towards building a synchronized chemotactic oscillator
- The assembly process, device characterization, and standard operating conditions are key enablers which will greatly benefit the field of synthetic biology



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